

U.S. Serial No. 08/07/2003

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Amendments to the Specification

At page 1, please amend the section entitled "Cross-reference to Related Applications" as follows:

This application is a continuation of U.S. Serial No. 09/787,092, filed March 12, 2001, now abandoned, which is a 371 National Stage application of PCT/US99/21731, filed March 12, 2001, and which is a continuation-in-part of U.S. Serial No. 09/158,406, filed September 22, 1998, which issued as U.S. Patent No. 6,495,333, the disclosures of which are incorporated herein by reference in their entirety.

At page 8, line 7, through page 9, line 27, please amend the section entitled "Brief Description of the Drawings" as follows:

The above and other objects and advantages of the invention will be apparent upon consideration of the following detailed description, taken in conjunction with the accompanying drawings, in which like reference characters refer to like parts throughout, and in which:

FIG. 1 is a flow chart schematizing the basic steps in a whole blood flow cytometric assay for dendritic cell function, with LPS exemplified as the dendritic cell activator;

FIGS. 2A-2I presents a series of dot plots generated during the flow cytometric analysis of whole blood activated with LPS in the presence of Brefeldin A, as described in example 3 - ~~CD11c⁺ dendritic cells are painted green and CD11c⁻ dendritic cells are painted red; nondendritic cells appear gray. The colors are arbitrarily chosen, and bear no relationship to the fluorochromes used for analysis;~~

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FIGS. 3A-3I presents a series of dot plots generated during the flow cytometric analysis of whole blood activated with PMA+I in the presence of Brefeldin A, as described in example 3. ~~CD11c⁺ dendritic cells are painted green and CD11c⁻ dendritic cells are painted red; nondendritic cells appear gray. The colors are arbitrarily chosen, and bear no relationship to the fluorochromes used for analysis;~~

FIGS. 4A-4K presents a series of dot plots generated during the flow cytometric analysis of whole blood incubated in the presence of ~~Brefeldin~~ brefeldin A in the absence of activator (resting control), as described in example 3. ~~CD11c⁺ dendritic cells are painted green and CD11c⁻ dendritic cells are painted red; nondendritic cells appear gray. The colors are arbitrarily chosen, and bear no relationship to the fluorochromes used for analysis;~~

FIGS. 5 presents the differential expression of TNF α and IL-8 in CD11c⁺ dendritic cells from two donors each activated alternatively with LPS or PMA+I;

FIGS. 6A-6C is a series of histograms summarizing the effects of three different dendritic activators on the surface expression of the identified markers on peripheral blood dendritic cells in whole blood;

FIG. 7A-B shows a comparison of cytokine expression between monocytes (gray bars) and CD11c⁺ DCs (black bars) in activated whole blood, with FIG. 7A showing LPS + Brefeldin A-stimulated cells, and FIG. 7B showing PMA+I + Brefeldin A-stimulated cells; and

FIG. 8 shows kinetics of TNF α , IL-1 β , IL-6 and CD80 in LPS activated CD11c⁺ DCs. The time course of LPS incubation was 0 to 8 hours. Intracellular cytokine and CD80 expression is measured as PE mean fluorescence intensity (MFI).

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At page 31, lines 8-23, please amend the paragraph extending from lines 8 through 23 as follows:

FIGS. 2A-2C show the surface immunophenotypic characteristics of peripheral blood DC from a single LPS-activated whole blood sample. ~~CD11c⁺ dendritic cells are painted green, CD11c⁻ DC are painted red, and nondendritic cells appear gray. The colors are arbitrarily chosen for purposes of display, and bear no relationship to the fluorophores used for analysis.~~ FIG. 2A demonstrates that both dendritic cell subsets are lin 1 FITC^{dim} and HLA-DR^{bright}, in agreement with O'Doherty *et al.*, *Immunology* 82: 487-493 (1994); Olweus *et al.*, *Proc. Natl. Acad. Sci. USA* 94 (23): 12551-12556 (1997), with FIG. 2B further demonstrating that the two subsets have similar side scatter and forward scatter properties. FIG. 2C shows discrimination of the two subsets based on differential levels of CD11c expression.